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Predictability of polygenic risk score for progression to dementia and its interaction with *APOE* ε4 in mild cognitive impairment

Jung-Min Pyun¹, Young Ho Park^{2*}, Keon-Joo Lee², SangYun Kim², Andrew J. Saykin^{3,4} and Kwangsik Nho^{3,5*} for the Alzheimer's Disease Neuroimaging Initiative

Abstract

Background: The combinatorial effect of multiple genetic factors calculated as a polygenic risk score (PRS) has been studied to predict disease progression to Alzheimer's disease (AD) from mild cognitive impairment (MCI). Previous studies have investigated the performance of PRS in the prediction of disease progression to AD by including and excluding single nucleotide polymorphisms within the region surrounding the *APOE* gene. These studies may have missed the *APOE* genotype-specific predictability of PRS for disease progression to AD.

Methods: We analyzed 732 MCI from the Alzheimer's Disease Neuroimaging Initiative cohort, including those who progressed to AD within 5 years post-baseline (n = 270) and remained stable as MCI (n = 462). The predictability of PRS including and excluding the *APOE* region (PRS_{+APOE} and PRS_{-APOE}) on the conversion to AD and its interaction with the *APOE* ϵ 4 carrier status were assessed using Cox regression analyses.

Results: PRS_{+APOE} (hazard ratio [HR] 1.468, 95% CI 1.335–1.615) and PRS_{-APOE} (HR 1.293, 95% CI 1.157–1.445) were both associated with a significantly increased risk of MCI progression to dementia. The interaction between PRS_{+APOE} and *APOE* ϵ 4 carrier status was significant with a *P*-value of 0.0378. The association of PRSs with the progression risk was stronger in *APOE* ϵ 4 non-carriers (PRS_{+APOE}⁻ HR 1.710, 95% CI 1.244–2.351; PRS_{-APOE}⁻ HR 1.429, 95% CI 1.182–1.728) than in *APOE* ϵ 4 carriers (PRS_{+APOE}⁻ HR 1.167, 95% CI 1.005–1.355; PRS_{-APOE}⁻ HR 1.172, 95% CI 1.020–1.346).

Conclusions: PRS could predict the conversion of MCI to dementia with a stronger association in APOE ϵ 4 noncarriers than APOE ϵ 4 carriers. This indicates PRS as a potential genetic predictor particularly for MCI with no APOE ϵ 4 alleles.

Keywords: Polygenic risk score, Mild cognitive impairment, Alzheimer's disease, Disease progression, APOE E4

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Background

Predicting disease progression to Alzheimer's disease (AD) from mild cognitive impairment (MCI) is critical for identifying individuals for opportune intervention in clinical management and for optimization of target participants in clinical trials. The use of individualized genetic profile is thriving with a pursuit of precision medicine and is becoming approachable with shared large data, which may facilitate customized risk prediction of disease development and progression. The heritability of

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late-onset AD (LOAD) is high, accounting for 60%–80% [1], and genome-wide association studies (GWAS) have identified multiple genetic factors associated with AD including the well-known *APOE* ε 4 allele [2, 3]. The combinatorial effect of these multiple genetic factors can be calculated as a polygenic risk score (PRS). PRS has been widely evaluated in AD research regarding its value in AD risk prediction, its relations with conventional biomarkers of AD, and its prediction of disease conversion from MCI to AD [4]. In particular, the predictability of PRS on disease progression from MCI to AD is substantially affected by the presence of *APOE* ε 4 allele, and PRS excluding the *APOE* region may or may not predict disease progression [5–8].

In this study, we aimed to investigate the predictability of PRSs including and excluding single nucleotide polymorphisms (SNPs) within the region surrounding the *APOE* gene, on MCI progression to AD, and the interaction between PRS and *APOE* ε 4 alleles. In addition, the predictability of hippocampal volume on MRI and amyloid PET were assessed, which are well-known predictors of disease progression. Furthermore, we identified SNPs used for PRS calculation and then performed enrichment analysis to explore the implicated biological pathways.

Methods

Participants

Data used in this study were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (http://adni.loni.usc.edu/). ADNI was launched in 2003 as a public-private partnership. The primary goal of ADNI has been to test whether serial MRI, PET, other biological markers, and clinical neuropsychological assessment can be combined to measure the progression of MCI and early AD [9]. In this study, patients with a diagnosis of MCI and with available GWAS data, older than 60 years at baseline assessment, and having at least one or more follow-up visits, were included. We focused on the genetic risk estimation of LOAD, in which the threshold cutoff of 60 years is commonly used. The primary outcome of this study was MCI conversion to dementia during a follow-up period up to 5 years postbaseline. MCI was diagnosed when there was objective memory impairment but without meeting the criteria for dementia [9]. The MCI participants had Mini-Mental State Examination (MMSE) scores between 24 and 30, memory performance scores approximately 1 standard deviation below expected education-adjusted norms, and a clinical dementia rating score of 0.5.

Genotyping and imputation

Genotyping for ADNI was performed using blood DNA samples and a combination of Illumina GWAS array

platforms (Illumina Human610-Quad BeadChip, Illumina HumanOmni Express BeadChip, and Illumina HumanOmni 2.5 M BeadChip) [10]. APOE genotyping was separately conducted using previously described standard methods to yield the APOE ɛ4 allele-defining SNPs (rs429358, rs7412) [10, 11]. Using PLINK 1.9 (www. cog-genomics.org/plink2/) [12], we performed standard quality control (QC) procedures for samples and SNPs as described previously [13]: (1) for SNPs: SNP call rate < 95%, Hardy–Weinberg *P* value < 1×10^{-6} , and minor allele frequency (MAF) < 1%; (2) for samples: sex inconsistencies, and sample call rate < 95%. Then, to prevent spurious associations due to population stratification, we used multidimensional scaling analysis to select only non-Hispanic participants of European ancestry that clustered with HapMap CEU (Utah residents with Northern and Western European ancestry from the CEPH collection) or TSI (Toscani in Italia) populations (Additional file 1: Fig. S1) [14, 15]. After QC procedures, we imputed un-genotyped SNPs separately in each platform using MaCH with the Haplotype Reference Consortium data as a reference panel [16, 17]. Following the imputation, we imposed an r^2 value of 0.30 as the threshold to accept the imputed genotypes [18].

Imaging biomarkers

T1-weighted brain MRI scan was processed with Free-Surfer version 5.1 to measure hippocampal and intracranial volumes [19]. For assessment of cortical amyloid accumulation, we used preprocessed (coregistered, averaged, standardized image and voxel size, uniform resolution) [¹⁸F] florbetapir PET scans [20] and calculated the mean standardized uptake value ratio (SUVR) using a whole cerebellum reference region as previously described [21]. In our analysis, amyloid burden on PET was dichotomized as positive when SUVR \geq 1.17 and negative when SUVR < 1.17 [22].

Calculation of PRS

PRSs were calculated using the software PRSice v2.3.1.e [23]. The GWAS summary statistics from Jansen et al. were used as a base dataset [2] and the phase 3 genetic data from the 1000 Genomes Project [24] for non-Hispanic participants of European ancestry were used to calculate the linkage disequilibrium structure. To investigate the optimal *P*-value threshold to select AD-associated SNPs, we iterated *P*-value thresholds of 1×10^{-6} , 1×10^{-5} , 1×10^{-4} , 1×10^{-3} , 1×10^{-2} , 0.05, and 0.5. PRSs were separately calculated including and excluding SNPs within the 1 Mb-region surrounding the *APOE* gene, which was defined as a region from 1 Mb before rs429358 to 1 Mb after rs7412. The numbers of SNPs used for PRS calculation depending on the various *P*-value thresholds

are shown in Additional file 1: Table S1. In addition, PRS was z-transformed based on the PRS distribution among amyloid PET-negative cognitively normal participants (n = 138).

Statistical analysis

All statistical analyses were made using the R v4.0.2 software (www.R-project.org). Comparisons of demographics between stable MCI, who remained stable at MCI during 5 years after baseline, and progressive MCI, who progressed to dementia within 5 years, were made with Mann–Whitney U test and Chi-squared test as appropriate.

To assess the predictability of PRS, hazard ratio (HR) of PRS in z-scores was obtained with Cox regression analysis. PRSs including and excluding SNPs within the 1 Mb-region surrounding the APOE gene were analyzed and represented as PRS_{+APOE} and PRS_{-APOE} , respectively. PRS analyses were performed in all MCI patients, MCI carrying the APOE ɛ4 alleles, and MCI not carrying the APOE ε 4 alleles, separately. Cox regression analyses of PRS are presented as Kaplan Meier curves. The interaction between PRSs and APOE £4 carrier status was also assessed. To evaluate the predictability of MRI hippocampal volume, the hippocampal volume was divided by the intracranial volume and z-transformed based on the distribution of values within amyloid PET-negative cognitively normal older adults. The HR of APOE £4 carriers was compared to APOE ɛ4 non-carriers, and the HR of amyloid PET-positive MCI was compared to amyloid PET-negative MCI. All analyses were adjusted with age and sex, and additionally with MRI field strength in hippocampal volume analysis. Statistical significance was set at *P* < 0.05.

Functional interpretation of SNPs used for PRS calculation

Functional mapping and annotation of SNPs used for PRS calculation was performed with the FUMA v1.3.6a software [25]. The GWAS summary statistics of SNPs were obtained from Jansen et al. [2]. All known SNPs that had $r^2 \ge 0.6$ with one of the independent significant SNPs used for PRS calculation, were also included for gene mapping using FUMA. Gene mapping was conducted by three methods, including positional mapping, expression quantitative trait loci (eOTL) mapping, and chromatin interaction mapping. In the positional mapping, SNPs were mapped to genes with a maximum distance of 10 kb. eQTL data from the eQTL catalogue (BrainSeq brain) [26], the Blood eQTLs (Westra et al. (2013) Blood eQTL Browser [27], Zhernakiva et al. (2017) BIOS QTL Browser [28]), the BRAINEAC (averaged expression of 10 brain regions including frontal cortex, hippocampus, occipital cortex, temporal cortex, cerebellar cortex, inferior olivary nucleus, putamen, substantia nigra, thalamus, and intralobular white matter) [29], and the GTEx v8 Brain (cortex, frontal cortex BA9, hippocampus)[30] datasets were used for eQTL mapping, and only significant SNP-gene pairs with a false discovery rate (FDR) cutoff of 0.05 were used. The chromatin interaction mapping was performed with data from the HiC (GSE87112) dorsolateral prefrontal cortex and hippocampus [31] with FDR cutoff of 1×10^{-6} and promotor region window of 250 bp up- and 500 bp down-stream of the transcription start site. The possible biological processes (BP), molecular functions (MF), and involved cellular components (CC) of mapped genes were explored by gene set enrichment analysis using data from the gene ontology (GO) [32, 33]. Statistical significance was set at adjusted *P*-value < 0.05.

Results

A total of 907 MCI participants from the ANDI cohort were assessed for study eligibility. After excluding 48 participants younger than 60 years, 68 without followup visit, and 59 without available GWAS data, 732 participants were finally included for the analysis. Of the 732 MCI participants, 270 MCI patients (36.8%) were converted to dementia within 5 years ("progressive MCI") and 462 MCI patients remained at MCI ("stable MCI"). There was no significant difference in age, sex, or education level at baseline between the two groups. The progressive MCI group had a significantly higher proportion of *APOE* ϵ 4 allele-carriers, lower MMSE scores and higher clinical dementia rating sum of boxes (CDR-SB) scores at baseline (Table 1).

We examined seven *P*-value thresholds to find the optimal threshold for selection of AD-associated SNPs, and the model with threshold $P < 1 \times 10^{-5}$ showed the best predictability and was used for further analysis (Additional file 1: Table S2).

In the Cox regression analysis to assess the predictability for the conversion of MCI to AD, PRS_{+APOE} (HR

 Table 1
 Baseline demographics of participants

	Stable MCI (n = 462)	Progressive MCI (n=270)	P-value
Age, years	73 (67–79)	74 (69–79)	0.109
Female, <i>n</i> (%)	179 (38.74)	105 (38.89)	1.000
Education, years	16 (14–18)	16 (14–18)	0.481
APOE ε 4 carrier, n (%)*	196 (42.52)	182 (67.41)	< 0.001
MMSE	28 (27–29)	27 (26–28)	< 0.001
CDR-SB	1 (0.5–1.5)	2 (1.0–2.5)	< 0.001

Data are presented as median (interquartile range) unless otherwise specified * Data unavailable for 1 subject

1.468, 95% CI 1.335–1.615) and PRS_APOF (HR 1.293, 95% CI 1.157-1.445) both presented a significant association with the risk of disease progression (Table 2). PRS_{+APOF} showed significantly increased HRs in both MCI with and without APOE ɛ4 alleles. In particular, the risk association in MCI with no APOE £4 alleles was stronger (HR 1.710, 95% CI 1.244-2.351) than MCI with APOE ε4 alleles (HR 1.167, 95% CI 1.005-1.355) (Table 2). The interaction between PRS_{+APOE} and APOE £4 carrier status in Cox regression analysis was also significant, with a coefficient of -0.372 (Table 3). PRS_{-APOE} also presented stronger risk association in MCI with no APOE ε 4 alleles (HR 1.429, 95% CI 1.182-1.728) than in MCI with APOE ε4 alleles (HR 1.172, 95% CI 1.020-1.346). However, the interaction between PRS_APOE and APOE £4 carrier status was not significant, with a coefficient of -0.174. HRs of PRS_{+APOE} and PRS_{-APOE} in MCI with and without APOE ε 4 alleles are presented with Kaplan Meier curves (Fig. 1).

Next, we performed Cox regression analysis to assess the predictability of AD-related biomarkers such as amyloid PET positivity and hippocampal volume on MRI, separately, for the conversion of MCI to AD, using imaging biomarkers as independent variables. The MCI participants carrying the *APOE* ε 4 allele showed increased HR (2.678, 95% CI 2.066–3.470) compared to the MCI not carrying the *APOE* ε 4 allele. The hippocampal volume on MRI was related to a decreased risk of disease progression (HR 0.563, 95% CI 0.502–0.632), and the amyloid PET positivity predicted disease progression with HR of 7.449 (95% CI 4.199–13.215) (Table 2).

For gene mapping, all SNPs (n = 4967) that had $r^2 \ge 0.6$ with one of the independent SNPs ($r^2 < 0.05$,

Table 3 Cox regression analysis of interaction between PRS and APOE ϵ 4 carrier status in association with disease progression from MCI to AD

	Coefficient	Standard error	P-value
Age	0.034	0.009	1.94×10^{-4}
Female	0.116	0.127	3.57×10^{-1}
PRS _{+APOE}	0.531	0.162	1.10×10^{-3}
APOE ɛ4 carrier status	0.681	0.176	1.14×10^{-4}
PRS _{+APOE} × APOE ε4 carrier status	- 0.372	0.179	3.78 × 10 ⁻²

 $\mathsf{PRS}_{+\textit{APOE}}$ PRS including SNPs within the 1 Mb-region surrounding the <code>APOE</code> gene

n=204) used to calculate PRS at a selection threshold of $P < 1 \times 10^{-5}$, were mapped on 424 genes. Among the 4967 SNPs, 3551 SNPs were used for positional mapping of 316 genes, 2898 SNPs for eQTL mapping of 264 genes, and 73 SNPs for chromatin interaction mapping of 19 genes. Among a total of 424 mapped genes, 133 genes were mapped by multiple independent significant SNPs. The mapped genes are listed in Additional file 2: Table S3. Gene set enrichment analysis yielded 16 significant GO BP pathways, 11 significant GO CC pathways, and no significant GO MF pathways (Fig. 2).

Discussion

In this study, we demonstrated that PRS_{+APOE} was significantly associated with an increased risk of progression to dementia in MCI. PRS_{-APOE} was also related to an increased risk of progression, albeit less pronounced. Notably, the association of PRS with progression risk was

Table 2 Association of PRS, APOE ε4 status, hippocampal volume on MRI, and amyloid PET positivity with disease progression to dementia according to z-scores in all MCI participants, MCI with APOE ε4, and MCI without APOE ε4

	HR (95% Cl), <i>P</i> -value			
	All MCI	MCl with <i>APOE</i> ε4	MCI without <i>APOE</i> ε4	
	(n=732)	(<i>n</i> = 378)	(<i>n</i> = 353)	
PRS _{+APOE}	1.468 (1.335–1.615)	1.167 (1.005–1.355)	1.710 (1.244–2.351)	
	2.30 × 10 ⁻¹⁵	4.16 × 10 ⁻²	9.30 × 10 ⁻⁴	
PRS_APOE	1.293 (1.157–1.445)	1.172 (1.020–1.346)	1.429 (1.182–1.728)	
	5.19 × 10 ⁻⁶	2.47 × 10 ⁻²	2.19 × 10 ⁻⁴	
APOE ɛ4 carrier status	2.678 (2.066–3.470) 9.70 × 10 ^{-14†}	NA	NA	
Hippocampal volume on MRI	0.563 (0.502–0.632)	0.538 (0.461–0.628)	0.536 (0.435–0.660)	
	<2.00 × 10 ^{-16*}	3.55 × 10 ^{–15†}	4.44 × 10 ^{-9††}	
Amyloid PET positivity	7.449 (4.199–13.215)	5.011 (1.966–12.764)	6.500 (2.914–14.495)	
	6.61 × 10 ⁻¹²⁵	7.31 × 10 ^{-4∥}	4.77 × 10 ^{-6¶}	

 PRS_{+APOE} : PRS including SNPs within the 1 Mb-region surrounding the *APOE* gene; PRS__{APOE}: PRS excluding SNPs within the 1 Mb-region surrounding the *APOE* gene Cox regression analysis of all variables were adjusted with age and sex, and additionally with MRI field strength in hippocampal volume analysis Data unavailable for * 3 subjects, † 1 subject, ⁵ 343 subjects, ⁸ 191 subjects, and ⁹ 152 subjects



stronger in *APOE* ε 4 non-carriers than in *APOE* ε 4 carriers in MCI, showing a significant interaction between PRS_{+APOE} and *APOE* ε 4 carrier status on the conversion of MCI to dementia.

A previous study showed that the higher PRS, excluding chromosome 19 SNPs to avoid the *APOE* effect, could predict clinical progression to MCI/AD from nondemented status, with unstandardized β value of 0.49 in logistic regression analysis. PRS including chromosome 19 has also shown similar results with unstandardized β value of 0.43 [5]. In another study, while PRS with *APOE* is significantly associated with the progression risk from MCI to AD with HR of 1.59 (95% CI 1.31–1.78), PRS without *APOE* could not predict the progression with HR of 1.03 (95% CI 0.79–1.34) [6]. In the current study, we found comparable HRs for PRS with and without *APOE* and additionally confirmed different predictability between *APOE* ε 4 carriers and non-carriers of the MCI participants.



Here, PRS_{-APOE} showed less prominent association with disease progression risk than PRS_{+APOE} which suggests that APOE is the strongest known risk gene for LOAD and affects the predictability of PRS. The APOE ε4 carriers show a three- to four-fold increase of risk of AD development compared to the APOE E4 non-carriers [34]. In addition to the effect of APOE alone, other ADassociated genes can interact with APOE to affect the disease course. A previous study has revealed that the SNP variants at FYN and RNF219 loci are associated with decreased LOAD age-of-onset in APOE £4 noncarriers but not in APOE ɛ4 carriers [35]. The RNF219 variant is also related with the beta-amyloid (Aß) load in APOE £4 non-carriers, but not in APOE £4 carriers [35]. Alleles of the *CETP* gene could alter AD risk in an APOE-dependent manner [36]. In a meta-analysis, CLU is associated with AD only in APOE £4 non-carriers and PICALM only in APOE £4 carriers [37]. The SNPs used for PRS calculation in our study also included SNPs of CLU and PICALM, and the integrated effect of interaction between genes containing SNPs of PRS and APOE could manifest as a stronger progression risk in APOE £4 non-carriers of our MCI cohort.

How the AD-associated genes impact AD pathology is not completely elusive, although the exact mechanism of the well-known APOE £4 allele for amyloid clearance and aggregation and its association with AD-associated genes remain unclear [38]. Recent studies have found that the APOE £4 allele is associated with Aß elevation and accumulation on PET, whereas PRS is more associated with faster cognitive decline in amyloid-positive status [39, 40]. This implies that APOE ε 4 contributes to the initiation of amyloidopathy in an early stage, while other genetic variants are involved in disease progression. In addition, pathway analysis has shown that a large proportion of the AD-related pathways is associated with the APOE region. Furthermore, pathways such as the protein-lipid complex subunit organization and protein-lipid complex assembly have also been found to be involved in the AD risk independent of the APOE region [24, 39]. Again, these pathways were confirmed in our GO BP results. Another study investigating the contribution of five pathway-specific PRSs to AD risk has shown that the most involved pathway is Aß metabolism (29.6%) when the analysis included APOE variants, and immune response (45.5%) when the analysis excluded APOE variants [41].

However, the pathways related to *APOE* and PRS cannot be distinguished clearly, and a complex interaction of Aß, tau metabolism, cholesterol/lipid metabolism [42,

43], endosomal–lysosomal processing [44], neuroinflammation [45, 46], cerebrovascular integrity [47, 48], and susceptibility to infectious agents [49] should be considered. We also observed that the genes corresponding to the SNPs used for the PRS calculation in this study were involved in diverse pathogenesis of AD, such as cholesterol metabolism (*APOE, CLU, ABCA7*), tau toxicity (*BIN1, CD2AP, FERMT2, CASS4, PTK2B*), immune response (*CR1, CD33, MS4A, TREM2*), and endocytosis (*BIN1, PICALM, CD2AP, SPHA1, SORL1*) [3, 50].

Another finding in our study was that PRS_{+APOE} and PRS_{-APOE} had relatively lower predictability for the conversion of MCI to AD, compared to MRI and amyloid PET biomarkers, which implies a limitation of PRS use in disease stages when abnormalities on MRI and amyloid PET biomarkers have fully developed. However, PRS as a non-modifiable risk factor given at birth could offer early prediction, independent of the disease stage.

Limitations and strengths

There are a few limitations in the current study. The sample size was small, so replication in an independent cohort would be desirable. With a longer follow-up period, some stable MCI participants might convert to AD. There are several challenges for PRS utility in clinical practice. Although the predictive utility of PRS has been validated in clinically well-defined and biomarkerconfirmed cohorts, the PRS risk estimation in clinical populations from the community could be affected by variables such as ethnicity, environmental factors, and mixed pathologies. In addition, PRS as an additive indicator should be interpreted with caution to avoid ethical issues such as genetic determinism [51, 52]. Furthermore, bioinformatics tools for fast and simple data processing are needed for clinical PRS application in patients. With these issues being solved, PRS would be a useful biomarker based on the important genetic contributions in diseases and improve the cost and accessibility.

PRS is also an emerging tool for genetic risk estimation in AD. The predictability of PRS for disease progression has been reported with inconsistency depending on the inclusion or exclusion of *APOE* ε 4 [4]. In this study, we not only investigated the predictability of PRS with and without *APOE* ε 4, but also analyzed, for the first time, the interaction between PRS and *APOE* ε 4. In addition, we present, for the first time, a comparison of the predictability of PRS for disease progression, with conventional imaging AD biomarkers including PET-based amyloid deposition and MRI-based brain atrophy.

Conclusions

In summary, we demonstrated that PRS could predict conversion of MCI to dementia, showing a significant interaction between PRS and *APOE* ε 4 carrier status, particularly stronger association in *APOE* ε 4 non-carriers than in carriers. These results support that PRS can predict disease progression in MCI patients with no *APOE* ε 4 allele, which has so far been the primary genetic risk factor used for clinical assessment. Furthermore, individualized risk estimation with PRS may allow timely interventions such as life style modification for disease prevention, which may reduce the dementia risk despite high genetic risk [53].

Abbreviations

Aß: Beta-amyloid; AD: Alzheimer's disease; ADNI: Alzheimer's Disease Neuroimaging Initiative; BP: Biological processes; CC: Cellular components; CDR-SB: Clinical dementia rating sum of boxes; CI: Confidence interval; eQTL: Expression quantitative trait loci; GO: Gene ontology; GWAS: Genome-wide association studies; HR: Hazard ratio; LOAD: Late-onset Alzheimer's disease; MAF: Minor allele frequency; MCI: Mild cognitive impairment; MF: Molecular functions; MMSE: Mini-Mental State Examination; PRS: Polygenic risk score; QC: Quality control; SNP: Single nucleotide polymorphism; SUVR: Standardized uptake value ratio.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s40035-021-00259-w.

Additional file 1: Fig. S1. The multidimensional scaling analysis for study population selection. Table S1. Number of SNPs for PRS calculation depending on GWAS association *P*-value thresholds. Table S2. Predictability of PRS for progression from mild cognitive impairment to dementia depending on GWAS *P*-value thresholds.

Additional file 2: Table S3. List of mapped genes.

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Authors' contributions

JMP: Analysis and interpretation of data, manuscript draft. YHP: Study design, analysis and interpretation of data, manuscript revision. SYK: Manuscript revision. KJL: Analysis and interpretation of data, manuscript revision. AJS: Interpretation of data, manuscript revision. KN: Study design, analysis and interpretation of data, manuscript revision.

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Availability of data and materials

All demographic, imaging, and genetic data in this study are publicly available and can be downloaded from the ADNI database (adni.loni.usc.edu).

Declarations

Ethics approval and consent to participate

The study procedures were approved by the institutional review board of all participating centres (http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf), and written informed consent was obtained from all participants or authorized representatives.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Gatz M, Reynolds CA, Fratiglioni L, Johansson B, Mortimer JA, Berg S, et al. Role of genes and environments for explaining Alzheimer disease. Arch Gen Psychiatry. 2006;63:168–74.
- 2. Jansen IE, Savage JE, Watanabe K, Bryois J, Williams DM, Steinberg S, et al. Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk. Nat Genet. 2019;51:404–13.
- Kunkle BW, Grenier-Boley B, Sims R, Bis JC, Damotte V, Naj AC, et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Aβ, tau, immunity and lipid processing. Nat Genet. 2019;51:414–30.
- Harrison JR, Mistry S, Muskett N, Escott-Price V. From polygenic scores to precision medicine in Alzheimer's disease: a systematic review. J Alzheimer's Dis. 2020;74:1271–83.
- Mormino EC, Sperling RA, Holmes AJ, Buckner RL, De Jager PL, Smoller JW, et al. Polygenic risk of Alzheimer disease is associated with early- and late-life processes. Neurology. 2016;87:481–8.
- Adams HHH, De Bruijn RFAG, Hofman A, Uitterlinden AG, van Duijn CM, Vernooij MW, et al. Genetic risk of neurodegenerative diseases is associated with mild cognitive impairment and conversion to dementia. Alzheimers Dement. 2015;11:1277–85.
- Lacour A, Espinosa A, Louwersheimer E, Heilmann S, Hernández I, Wolfsgruber S, et al. Genome-wide significant risk factors for Alzheimer's disease: role in progression to dementia due to Alzheimer's disease among subjects with mild cognitive impairment. Mol Psychiatry. 2017;22:153–60.
- Carrasquillo MM, Crook JE, Pedraza O, Thomas CS, Pankratz VS, Allen M, et al. Late-onset Alzheimer's risk variants in memory decline, incident mild cognitive impairment, and Alzheimer's disease. Neurobiol Aging. 2015;36:60–7.
- Aisen PS, Petersen RC, Donohue M, Weiner MW. Alzheimer's Disease Neuroimaging Initiative 2 Clinical Core: progress and plans. Alzheimer's Dement. 2015;11:734–9.
- Saykin AJ, Shen L, Yao X, Kim S, Nho K, Risacher SL, et al. Genetic studies of quantitative MCI and AD phenotypes in ADNI: Progress, opportunities, and plans. Alzheimers Dement. 2015;11:792–814.
- Furney SJ, Simmons A, Breen G, Pedroso I, Lunnon K, Proitsi P, et al. Genome-wide association with MRI atrophy measures as a quantitative trait locus for Alzheimer's disease. Mol Psychiatry. 2011;16:1130–8.
- Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet. 2007;81:559–75.
- Lee Y, Han S, Kim D, Kim D, Horgousluoglu E, Risacher SL, et al. Genetic variation affecting exon skipping contributes to brain structural atrophy in Alzheimer's disease. AMIA Jt Summits Transl Sci Proc. 2018;2017:124–31.
- Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D. Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet. 2006;38:904–9.
- 15. Thorisson GA, Smith AV, Krishnan L, Stein LD. The international HapMap project web site. Genome Res. 2005;15:1592–3.
- Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: Using sequence and genotype data to estimate haplotypes and unobserved genotypes. Genet Epidemiol. 2010;34:816–34.
- McCarthy S, Das S, Kretzschmar W, Delaneau O, Wood AR, Teumer A, et al. A reference panel of 64,976 haplotypes for genotype imputation. Nat Genet. 2016;48:1279–83.
- Park YH, Hodges A, Simmons A, Lovestone S, Weiner MW, Kim S, et al. Association of blood-based transcriptional risk scores with biomarkers for Alzheimer disease. Neurol Genet. 2020;6:e517.
- Jack CR Jr, Bernstein AMA, Borowski ABJ, Gunter JL, Fox NC, Thompson PM, et al. Update on the MRI core of the Alzheimer's Disease Neuroimaging Initiative. Alzheimers Dement. 2010;6:212–20.

- Jagust WJ, Landau SM, Koeppe RA, Reiman EM, Chen K, Mathis CA, et al. The Alzheimer's Disease Neuroimaging Initiative 2 PET Core: 2015. Alzheimers Dement. 2015;11:757–71.
- Risacher SL, Kim S, Nho K, Foroud T, Shen L, Petersen RC, et al. APOE effect on Alzheimer's disease biomarkers in older adults with significant memory concern. Alzheimers Dement. 2015;11:1417–29.
- Fleisher AS, Chen K, Liu X, Roontiva A, Thiyyagura P, Ayutyanont N, et al. Using positron emission tomography and florbetapir F18 to image cortical amyloid in patients with mild cognitive impairment or dementia due to Alzheimer disease. Arch Neurol. 2011;68(11):1404–11.
- Euesden J, Lewis CM, O'Reilly PF. PRSice: Polygenic Risk Score software. Bioinformatics. 2015;31:1466–8.
- Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, et al. A global reference for human genetic variation. Nature. 2015;526:68–74.
- Watanabe K, Taskesen E, Van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. Nat Commun. 2017;8:1–10.
- Kerimov N, Hayhurst JD, Manning JR, Walter P, Kolberg L, Peikova K, et al. eQTL catalogue: a compendium of uniformly processed human gene expression and splicing QTLs. bioRxiv. 2020; preprint: not peer reviewed. https://doi.org/10.1101/2020.01.29.924266.
- Westra HJ, Peters MJ, Esko T, Yaghootkar H, Schurmann C, Kettunen J, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. Nat Genet. 2013;45:1238–43.
- Zhernakova DV, Deelen P, Vermaat M, van Iterson M, van Galen M, Arindrarto W, et al. Identification of context-dependent expression quantitative trait loci in whole blood. Nat Genet. 2017;49:139–45.
- Ramasamy A, Trabzuni D, Guelfi S, Varghese V, Smith C, Walker R, et al. Genetic variability in the regulation of gene expression in ten regions of the human brain. Nat Neurosci. 2014;17:1418–28.
- Aguet F, Barbeira AN, Bonazzola R, Gabriel S, Getz G, Graubert A, et al. The GTEx Consortium atlas of genetic regulatory effects across human tissues The Genotype Tissue Expression Consortium. Science. 2020;369(6509):1318–30.
- Schmitt AD, Hu M, Jung I, Xu Z, Qiu Y, Tan CL, et al. A compendium of chromatin contact maps reveals spatially active regions in the human genome. Cell Rep. 2016;17:2042–59.
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. Nat Genet. 2000;25:25–9.
- The Gene Ontology Consortium. The Gene Ontology Resource: 20 years and still GOing strong. Nucleic Acids Res. 2019;47(D1):D330–8.
- Yamazaki Y, Zhao N, Caulfield TR, Liu CC, Bu G. Apolipoprotein E and Alzheimer disease: pathobiology and targeting strategies. Nat Rev Neurol. 2019;15:501–18.
- Rhinn H, Fujita R, Qiang L, Cheng R, Lee JH, Abeliovich A. Integrative genomics identifies APOE ε4 effectors in Alzheimer's disease. Nature. 2013;500:45–50.
- Murphy EA, Roddey JC, Mcevoy LK, Holland D, Hagler DJ Jr, Dale AM, et al. CETP polymorphisms associate with brain atrophy rate, and Alzheimer's disease risk in an APOE-dependent manner. Brain Imaging Behav. 2015;6:16–26.

- Jun G, Naj AC, Beecham GW, Wang LS, Buros J, Gallins PJ, et al. Metaanalysis confirms CR1, CLU, and PICALM as Alzheimer disease risk loci and reveals interactions with APOE genotypes. Arch Neurol. 2010;67:1473–84.
- Belloy ME, Napolioni V, Greicius MD. A quarter century of APOE and Alzheimer's disease: progress to date and the path forward. Neuron. 2019;101:820–38.
- Leonenko G, Shoai M, Bellou E, Sims R, Williams J, Hardy J, et al. Genetic risk for Alzheimer disease is distinct from genetic risk for amyloid deposition. Ann Neurol. 2019;86:427–35.
- Ge T, Sabuncu MR, Smoller JW, Sperling RA, Mormino EC. Dissociable influences of APOE ε4 and polygenic risk of AD dementia on amyloid and cognition. Neurology. 2018;90:e1605–12.
- Tesi N, van der Lee SJ, Hulsman M, Jansen IE, Stringa N, van Schoor NM, et al. Immune response and endocytosis pathways are associated with the resilience against Alzheimer's disease. Transl Psychiatry. 2020;10:332.
- 42. Hamanaka H, Katoh-Fukui Y, Suzuki K, Kobayashi M, Suzuki R, Motegi Y, et al. Altered cholesterol metabolism in human apolipoprotein E4 knockin mice. Hum Mol Genet. 2000;9:353–61.
- Gong JS, Morita SY, Kobayashi M, Handa T, Fujita SC, Yanagisawa K, et al. Novel action of apolipoprotein e (ApoE): ApoE isoform specifically inhibits lipid-particle-mediated cholesterol release from neurons. Mol Neurodegener. 2007;2:9.
- 44. Nuriel T, Peng KY, Ashok A, Dillman AA, Figueroa HY, Apuzzo J, et al. The endosomal–lysosomal pathway is dysregulated by APOE4 expression in vivo. Front Neurosci. 2017;11:1–12.
- 45. Rodriguez GA, Tai LM, LaDu MJ, Rebeck GW. Human APOE4 increases microglia reactivity at A β plaques in a mouse model of A β deposition. J Neuroinflammation. 2014;11:111.
- Shi Y, Yamada K, Liddelow SA, Smith ST, Zhao L, Luo W, et al. ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. Nature. 2017;549:523–7.
- Tai LM, Thomas R, Marottoli FM, Koster KP, Kanekiyo T, Morris AW, et al. The role of APOE in cerebrovascular dysfunction. Acta Neuropathol. 2016;176:139–48.
- Montagne A, Nation DA, Sagare AP, Barisano G, Sweeney MD, Chakhoyan A, et al. APOE4 leads to blood–brain barrier dysfunction predicting cognitive decline. Nature. 2020;581:71–6.
- Linard M, Letenneur L, Garrigue I, Doize A, Dartigues JF, Helmer C. Interaction between APOE4 and herpes simplex virus type 1 in Alzheimer's disease. Alzheimers Dement. 2020;16:200–8.
- Karch CM, Goate AM. Alzheimer's disease risk genes and mechanisms of disease pathogenesis. Biol Psychiatry. 2015;77:43–51.
- Lewis CM, Vassos E. Polygenic risk scores: from research tools to clinical instruments. Genome Med. 2020;12(1):44.
- Torkamani A, Wineinger NE, Topol EJ. The personal and clinical utility of polygenic risk scores. Nat Rev Genet. 2018;19(9):581–90.
- Lourida I, Hannon E, Littlejohns TJ, Langa KM, Hyppönen E, Kuzma E, et al. Association of lifestyle and genetic risk with incidence of dementia. JAMA. 2019;322:430–7.

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