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One year brain atrophy evident in healthy aging

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Abstract

An accurate description of changes in the brain in healthy aging is needed to understand the basis of age-related changes in cognitive function. Cross-sectional magnetic resonance imaging (MRI) studies suggest thinning of the cerebral cortex, volumetric reductions of most subcortical structures and ventricular expansion. However, there is a paucity of detailed longitudinal studies to support the cross-sectional findings. In the present study, 142 healthy elderly participants (60–91 years) were followed with repeated MRI, and were compared to 122 patients with mild to moderate Alzheimer's disease (AD). Volume changes were measured across the entire cortex and in 48 regions of interest (ROIs). Cortical reductions in the healthy elderly were extensive after only one year, especially evident in temporal and prefrontal cortex where annual decline was about 0.5%. All subcortical and ventricular regions except caudate nucleus and the 4th ventricle changed significantly over one year. Some of the atrophy occurred in areas vulnerable to AD, while other changes were observed in areas less characteristic of the disease in early stages. This suggests that the changes are not primarily driven by degenerative processes associated with AD, although it is likely that preclinical changes associated with AD are superposed on changes due to normal aging in some subjects, especially in the temporal lobes. Finally, atrophy was found to accelerate with increasing age, and this was especially prominent in areas vulnerable to AD. Thus, it is possible that the accelerating atrophy with increasing age is due to preclinical AD.

Keywords

MRI; aging; longitudinal; ADNI; cerebral cortex; hippocampus

Introduction

An accurate description of age-related changes in the brain is needed to better understand the neurobiological foundation of cognitive changes in healthy aging. While cross-sectional magnetic resonance imaging (MRI) studies have demonstrated consistent age-effects (Allen et al., 2005; Fjell et al., 2009-a, b; Raz et al., 2004; Raz and Rodrigue, 2006; Salat et al., 2004; Walhovd et al., 2005; Westlye et al., 2009), longitudinal MR studies have often been limited by relatively small numbers of participants and use of either global measures (e.g. whole-brain volume) or only a restricted set of regions of interest (ROIs). Further, while atrophy has been demonstrated in mild cognitive impairment over a year or less (Schuff et al.,

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2009), it is not known whether brain changes driven solely by normal aging are large enough to be detectable over such short time intervals.

Longitudinal studies have reported annual gross brain volume decreases on the order of 0.2 -0.5% (Enzinger et al., 2005; Ezekiel et al., 2004; Fotenos et al., 2005; Scahill et al., 2003), while hippocampus, the most studied structure, shows annual atrophy rates from 0.79% to 2.0%(Barnes et al., 2009; Du et al., 2006; Ezekiel et al., 2004; Jack et al., 1998; Raz et al., 2005b; Scahill et al., 2003). This tends to be higher than cross-sectional estimates (Du et al., 2006; Raz et al., 2005a; Scahill et al., 2003). Entorhinal cortex longitudinal decline is about 0.3% to 2.4% (Du et al., 2006; Du et al., 2003; Ezekiel et al., 2004; Raz et al., 2005a). Atrophy rates in other parts of the brain are seldom studied, with a few important exceptions (Driscoll et al., 2009; Pfefferbaum et al., 1998; Raz et al., 2005a; Resnick et al., 2003). These studies describe atrophy in several regions, with prominent decline in prefrontal cortex, as well as decline in caudate nucleus, cerebellum, hippocampus and association cortices (Raz et al., 2005b) and parietal areas (Resnick et al., 2003), in addition to expansion of the ventricles (Pfefferbaum et al., 1998). In a recent study, significant atrophy was found in all 25 regional brain volumes tested, but over time periods extending as long as ten years in some cases (Driscoll et al., 2009). An earlier study found significant ventricular enlargement over one year, but no detectable change in total or regional brain volumes (Resnick et al., 2000).

With newer analyses techniques, we may now possess the tools necessary to detect brain changes over short time intervals in healthy elderly. The aims of the present study were to determine whether brain changes in normal aging can be detected over one (n = 142) or two (n = 89) years, and if so, to determine which areas show the greatest changes, and whether rate of atrophy increases with age. A novel procedure for longitudinal change analysis was used, which enabled measures of change continuously across the cortex, as well as in 48 regions of interest. Atrophy in the healthy elderly was compared to 122 patients with mild to moderate Alzheimer's disease (AD).

Materials and Methods

The raw data used in the preparation of this article were obtained from the ADNI database (www.loni.ucla.edu/ADNI). ADNI was launched in 2003 by the National Institute on Aging (NIA), the National Institute of Biomedical Imaging and Bioengineering (NIBIB), the Food and Drug Administration (FDA), private pharmaceutical companies and non-profit organizations. The primary goal of ADNI has been to test whether serial MRI, PET, other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and early AD. The Principal Investigator of this initiative is Michael W. Weiner, VA Medical Center and University of California — San Francisco. There are many co-investigators, and subjects have been recruited from over 50 sites across the U.S. and Canada. The initial goal of ADNI was to recruit 800 adults, including healthy elderly, MCI and AD patients to participate and be followed for 2–3 years. For more information see www.adni-info.org.

Sample

ADNI eligibility criteria are described at

http://www.adni-info.org/index.php?option=com_content&task=view&id=9&Itemid=43. Briefly, participants are 55–90 years of age, had an informant able to provide an independent evaluation of functioning, and spoke either English or Spanish. All subjects were willing and able to undergo test procedures including neuroimaging and agreed to longitudinal follow up. Specific psychoactive medications are excluded. General inclusion/exclusion criteria are as follows for normal subjects: Mini-Mental State Examination (MMSE) (Folstein et al., 1975) scores between 24–30 (inclusive), Clinical Dementia Rating (CDR) (Morris, 1993) of 0, non-

depressed, non-MCI, and nondemented. In addition, CDR sum of boxes (CDR-sb) was calculated as a measure of clinical functioning (does not need to be zero). To minimize the possibility of including individuals with early, preclinical AD, only participants who had the same or better score on CDR-sb at the time of follow up were included. The subject pool was further restricted to those subjects for whom adequate processed and quality checked MR data were available by February 2009. Sample characteristics are presented in Table 1.

To be able to compare atrophy in the healthy elderly with AD patients, an AD group was included for selected analyses (n = 122, 57 females/ 65 males, at 1 year follow up: age = 76.5 (56.0–90.0), MMSE = 20.7 (8–29), CDR = 1.0 (0.5–3.0), CDR-sb = 5.88 (1–14)). These patients met the same eligibility criteria as the healthy elderly, had CDR scores of 0.5 or more, and met NINCDS/ ADRDA criteria for probable AD (McKhann et al., 1984). As the mean age (76.5 vs. 76.7 years, respectively) and the female/ male ratio (0.88 vs. 0.92) was almost identical between AD patients and healthy elderly, these variables were not regressed out in the analyses.

MR acquisition and analysis

All scans used for the present paper were from 1.5 T scanners. Data were collected across a variety of scanners with protocols individualized for each scanner, as defined at http://www.loni.ucla.edu/ADNI/Research/Cores/index.shtml. Raw DICOM MRI scans (including two T1-weighted volumes per case) were downloaded from the public ADNI site (http://www.loni.ucla.edu/ADNI/Data/index.shtml) and processed as described elsewhere (Fennema-Notestine et al., 2009). Briefly, these data were reviewed for quality, automatically corrected for spatial distortion due to gradient nonlinearity (Jovicich et al., 2006) and B₁ field inhomogeneity (Sled et al., 1998), registered, and averaged to improve signal-to-noise. Scans were segmented as described previously (Fischl et al., 2002, 2004) to yielding data for 11 different volumetric ROIs and four ventricular ROIs (right and left hemisphere regions measured separately), including the hippocampal formation (consisting of the dentate gyrus, CA fields, subiculum/ parasubiculum and the fimbria (Makris et al., 1999)). The cortical surface was reconstructed using a semi-automated approach described elsewhere (Dale et al., 1999; Dale, 1993; Fischl and Dale, 2000; Fischl et al., 1999a; Fischl et al., 1999b; Salat et al., 2004), and parcellated as described in Fischl et al. (Fischl et al., 2004), labelling 33 cortical sulci and gyri (Desikan et al., 2006). The exact anatomical borders for the cortical ROIs can be found in Desikan et al., e.g. for the transverse temporal cortex (the rostral boundary was the rostral extent of the transverse temporal sulcus, the caudal boundary was the caudal portion of the insular cortex, and the lateral fissure and the superior temporal gyrus were utilized as the medial and lateral boundaries, respective) and isthmus of the cingulate cortex (the rostral and caudal boundaries were the posterior division of the cingulate cortex and the parahippocampal gyrus, respectively, and the medial and lateral boundaries were the medial wall (area unknown) and the precuneus, respectively). These surface-based cortical ROIs were thus generated by a different procedure from that used to estimate the other (volumetric) ROIs, which included the hippocampus, the cerebellum GM, nine subcortical ROIs and four ventricular ROIs.

For the analyses of the longitudinal volume changes, each participant's dual 3-D follow-up structural scans were rigid-body aligned, averaged and affine aligned to the baseline scan. Nonlinear registration of the images was then performed, where voxel centers are moved about until a good match between the images is made. Several methods exist for effecting this, including fluid deformation (Christensen et al., 1996; Freeborough and Fox, 1998; Miller et al., 1993) and tensor-based morphometry (TBM) (Ashburner et al., 1999). For the results presented here, however, we applied a method (Holland et al., 2008) based on linear elasticity and closer in spirit to TBM. It proceeds essentially as follows. The images are heavily blurred (smoothed), making them almost identical, and a merit or potential function calculated. This merit function expresses the intensity difference between the images at each voxel, and depends

on the displacement field for the voxel centers of the image being transformed; it is also regularized to keep the displacement field spatially smooth. The merit function by design will have a minimum when the displacement field induces a good match between the images. The displacement field in general will turn cubic voxels into displaced irregular hexahedra whose volumes (Grandy, 1997) give the volume change field. The merit function is minimized efficiently using standard numerical methods. Having found a displacement field for the heavily blurred pair of images, the blurring is reduced and the procedure repeated, thus iteratively building up a better displacement field. Two important additions to this are: applying the final displacement field to the image being transformed, then nonlinearly registering the resultant image to the same target, and finally tracing back through the displacement fields thus calculated to find the net displacement field; and restricting to regions of interest and zooming when tissue structures are separated by only a voxel or two. These additional features enable very precise registration involving large or subtle deformations, even at small spatial scales with low boundary contrast. Although large deformations are allowed by multiple nonlinear registration (or relaxation) steps, nonphysical deformations are precluded because at each level of blurring the image undergoing deformation is constrained to conform to the target. Note that calculating the deformation field does not depend on initially segmenting tissue. This deformation field was used to align scans at the sub-voxel level.

The follow-up aligned image underwent skull stripping and volumetric segmentations (subcortical structures, as well as hippocampus and cerebellum GM), with labels applied from the baseline scan. For the cortical reconstructions, surface coordinates for the white matter and pial boundaries were derived from the baseline images and mapped onto the follow-up images using the deformation field. Parcellations from the baseline image were then applied to the follow-up image. This resulted in a one-to-one correspondence between each vertex in the base image and the follow up image. The procedure produces an estimate of the percent cortical volume loss at each vertex and within each ROI. To the extent that regional cortical areas are relatively stable across time points, the volume change is likely driven almost exclusively by changes in thickness. The method has been validated in model studies of complex spherical-shell geometries with low contrast and noise, where a prescribed volume change is numerically estimated to accuracies of within 0.5% (Holland et al., work in progress).

A challenge in cross-sectional MR studies of aging is that the contrast between GM and WM decreases with higher age (Salat et al., 2009). However, the longitudinal registration relies on matching intensities in serial images. A reduction in GM/ WM contrast in elderly relative to younger participants should not impact the quality of the nonlinear registration, as long as the serial images have constant contrast. Change in cortical volume to 1st order arises from change in cortical thickness. To estimate it, the volume change field is sampled midway through the cortical layer. Conceivably, the primary source of error in elderly, relatively to younger participants, would then be in the initial delineation of the cortical surface. However, it has recently been shown that the decrease in GM/ WM contrast can not explain the differences in cortical thickness between older and younger participants (Westlye et al., 2009).

Statistical analyses

Statistical analyses were performed by use of FreeSurfer

(http://surfer.nmr.mgh.harvard.edu/) and SPSS 15.0. To reduce the number of comparisons, mean values of the left and right hemisphere were used for the ROI analyses. For the healthy elderly and the AD patients, change in brain morphometry was calculated as percentage change relative to baseline (1 and 2 years for the healthy elderly, 2 years for the AD patients). For the healthy elderly, statistical parametric continuous surface maps were computed by general linear models (GLM) implemented in FreeSurfer and displayed on a semi-inflated template brain. The results were thresholded by a conventional criterion for correction for multiple

comparisons (false discovery rate [FDR] < .05). For the healthy elderly and the AD patients, one-sample t-tests were used to test whether longitudinal atrophy rates within each ROI were different from zero. Independent-samples t-tests were used to compare rate of atrophy in healthy controls and AD patients. To visualize the areas that showed more than average and the areas that showed less than average atrophy as surface maps, mean cortical atrophy was calculated for each hemisphere in each group. This value was subtracted from the original surface maps, yielding a new map showing relative atrophy. To test whether the magnitude of atrophy in the temporal lobe vs. the prefrontal cortex differed between the healthy elderly and the AD patients, ANOVA with 2 groups \times 2 regions were performed. Temporal lobe atrophy was calculated as mean rate of atrophy in inferior and middle temporal, entorhinal, parahippocampal, temporal pole, and fusiform cortex, as well as hippocampus and amygdala. Prefrontal atrophy was calculated as mean rate of atrophy in caudal and rostral anterior cingulate, caudal and rostral middle frontal, pars opercularis, triangularis and orbitalis (together forming the inferior frontal cortex), lateral, dorsal and medial orbitofrontal, frontal pole and superior frontal cortex (see Figure 4). Finally, Pearson correlations were performed between age and atrophy in each of the 48 ROIs in the healthy elderly.

Results

Atrophy over 1 and 2 years: Continuous surface analyses

The statistical significance of 1 and 2 year volume changes in cortex across the brain surface is shown in Figure 1. As can be seen, large cortical areas decreased significantly over 1 year. These reductions were most prominent in the temporal lobe, the medial part of the superior frontal gyrus, orbitofrontal, precuneus, supramarginal and inferior parietal, as well as the inferior and part of the middle frontal cortex. Lateral parts of the superior frontal cortex, as well as pre- and post-central gyrus, superior parietal areas, occipital cortex and lingual gyrus were largely spared. Over two years, the effects were stronger and spread to additional areas, involving all cortical areas except lingual cortex and pre- and postcentral gyrus.

Percentage volume change in cortex over 1 and 2 years is shown in Figure 2. Annual atrophy rates varied across the cortex, but were typically around 0.5 % in affected areas, including most of the lateral and inferior parts of the temporal lobes, supramarginal gyrus, inferior parietal gyrus, precuneus, inferior and middle frontal gyrus as well as the medial parts of the superior frontal gyrus. Atrophy over 2 years exceeded 1.0 % in large areas.

For comparison purposes, rate of atrophy is presented for AD patients in Figure 2. Atrophy in the AD group was much larger than in the healthy elderly, with most areas showing 1% annual change or more. This was especially evident in the lateral temporal cortex and an area around posterior cingulate/ inferior precuneus/ retrosplenial cortex (> 2.5% annual change). Less atrophy was seen around the central sulcus, pericalcarine sulcus and lingual gyrus. To compare the pattern of atrophy between the healthy controls and the AD patients, mean cortical atrophy was calculated for each hemisphere in each group. This value was subtracted from the surface maps, yielding a new map highlighting areas that show greater and lesser rates of atrophy than average for that group. These maps illustrate the parts of the cortex that show relatively higher and relatively lower rates of atrophy within each group (Figure 3). Within the AD group, higher than average atrophy was seen in the inferior and middle temporal gyrus. Within the healthy elderly group, higher than average atrophy was observed in the same areas, but also in the temporal pole, medial orbitofrontal, rostral middle frontal and supramarginal cortex, as well as parts of the medial superior frontal gyrus. Less than average atrophy was seen in cuneus, pericalcarine sulcus, lingual gyrus as well as the central sulcus. Results of formal statistical tests of differences in rates of atrophy between healthy controls and AD patients are described below.

Atrophy over 1 and 2 years: ROI analyses

Percentage annual volume change in each of the 48 ROIs is shown in Table 2 for healthy elderly. Median atrophy for 1 year was -0.38 %, not including the ventricular ROIs. Hippocampus (-0.84 %) and amygdala (-0.81 %) showed the largest reductions, followed by thalamus (-0.69 %). Cerebellum grey matter (-0.35 %), the brain stem (-0.31 %) and caudate nucleus (-0.24 %) showed the least 1 year change of the volumetric ROIs. The inferior lateral (5.47 %), lateral (4.40 %) and the 3rd (3.07 %) ventricles showed the largest changes overall, with no significant change in the 4th (0.71 %) ventricle. Of the cortical ROIs, the frontal pole (-0.59 %), temporal pole (-0.56 %), banks of the superior temporal sulcus (-.55 %), and entorhinal cortex (-0.55 %) changed the most, while post central, pericalcarine, lingual, cuneus and precentral were well preserved (≤ 0.1 %).

For comparison purposes, 1 year volume change in each ROI was calculated for the AD group also (Table 3). All ROIs except the pericalcarine sulcus showed significant change. Median atrophy for 1 year was -1.20 %, not including the ventricular ROIs. Amygdala (-3.79 %) and hippocampus (-3.75 %) largest reductions, followed by inferior and middle temporal gyri (-2.97 and -2.87%, respectively). Also entorhinal and parahippocampal cortex showed well above average atrophy (-2.42 and -2.05%, respectively). Pericalcarine (-0.16%), postcentral (-0.27%) and cuneus (-0.5%) were the three ROIs showing the least atrophy, but of these only pericalcarine did not change at p < .01 (p = .030). The inferior lateral (16.26 %), lateral (10.35 %) and the 3rd (6.82 %) ventricles showed the largest changes overall.

Differences in rates of atrophy were formally tested between healthy elderly and AD patients. AD patients had significantly (p < .01) higher rates of change than the healthy elderly for all ROIs except 4th ventricle (Supplementary Table 1). In terms of t-scores, inferior temporal (t = 13.25), fusiform (t = 13.15) and middle temporal (t = 12.81), parahippocampal (t = 12.69) and entorhinal (t = 11.93) showed the largest effects of the cortical ROIs, while hippocampus (t = 12.80), the inferior lateral ventricles (t = 9.74) and amygdala (t = 8.32) showed the largest effects for the volumetric ROIs.

Interaction between the groups and temporal vs. frontal atrophy rates were tested by ANOVA with two groups (healthy elderly, AD) × two brain regions (frontal, temporal). A highly significant interaction was found (F [1, 256] = 119.78, p < 10^{-22}), caused by a higher rate of temporal than frontal atrophy in the AD group and a comparable rate of temporal vs. frontal atrophy rate in the healthy elderly (Figure 4).

Effects of age on regional atrophy rates

Correlation analyses between age and 1 year atrophy in each of the ROIs were performed. The results are presented in Table 4 and scatterplots for selected structures in Figure 5. We focus here on the 1 year changes, since the statistical power was lower for 2 years. Significant correlations between age and rates of atrophy were found for 32 ROIs. For the volumetric structures, hippocampus, cerebellum grey and white matter, cerebral white matter, thalamus and pallidum atrophy were related to age, as was expansion of the inferior lateral ventricles. Of the cortical structures, regions demonstrating significant relationships between age and rate of atrophy included entorhinal, transverse temporal, pars opercularis, isthmus cingulate, middle temporal, pars triangularis, banks of the superior temporal sulcus, parahippocampal, superior temporal, posterior cingulate, caudal middle frontal, precentral, fusiform, inferior parietal, temporal pole, precuneus, supramarginal, lateral orbitofrontal, superior parietal, post central, caudal anterior cingulate, medial orbitofrontal, pericalcarine, lateral occipital and inferior temporal cortices.

Discussion

Volumetric reductions of the cerebral cortex and subcortical brain structures, as well as expansion of the ventricles, were seen in healthy aging after only one year. While this has been shown previously for MCI and AD patients (McDonald et al., 2009; Schuff et al., 2009), the present report is the first to demonstrate detectable atrophy in a range of brain areas over only one year in healthy participants. Some of the changes seen, e.g. in the temporal lobe, may be due to preclinical AD, while others, e.g. in the prefrontal cortex, likely reflects the non-pathological aging process. This is discussed below.

The present atrophy rates for hippocampus (0.84%) and entorhinal cortex (0.55%) are comparable to previous studies (Barnes et al., 2009; Du et al., 2006; Du et al., 2003; Ezekiel et al., 2004; Jack et al., 1998; Raz et al., 2005a; Raz et al., 2005b; Scahill et al., 2003), supporting the feasibility of pooling data across multiple sites resulting in sensitive and realistic atrophy estimates. Generally, volumetric ROIs showed more decline than surface-based cortical ROIs, with the hippocampus showing the greatest one year reduction. Hippocampal volume has been found to correlate with age in cross-sectional (Good et al., 2001; Raz et al., 2004; Walhovd et al., 2005) and longitudinal studies (Driscoll et al., 2009; Du et al., 2006; Ezekiel et al., 2004; Jack et al., 1998; Raz et al., 2005b; Resnick et al., 2003; Scahill et al., 2003), but usually not more strongly than the other volumetric structures measured (Raz et al., 2005b). However, in a recent cross-sectional multi-sample study, estimated percentage decline per decade in the later half of the life-span was larger for hippocampus than any other volumetric structure (Walhovd et al., in press). In the present study, amygdala, thalamus, putamen and pallidum also showed longitudinal decline. Several cross-sectional studies have shown substantial reductions in the volumes of thalamus, amygdala and putamen (Greenberg et al., 2008; Gunning-Dixon et al., 1998; Raz et al., 2003; Sullivan et al., 2004; Van Der Werf et al., 2001; Walhovd et al., 2005; Walhovd et al., in press), as well as in pallidum after 50 years of age (Walhovd et al., 2005; Walhovd et al., in press), but discrepant findings regarding thalamus and pallidum have been reported (Gunning-Dixon et al., 1998; Jernigan et al., 1991; Jernigan et al., 2001; Luft et al., 1999; Raz et al., 2003).

Hippocampus and amygdala were the two volumetric structures that showed the largest annual decline in the present study, and these are known to show accelerated atrophy early in AD (Barnes et al., 2009; de Leon et al., 1989; Fennema-Notestine et al., 2009; Fischl et al., 2002; Jack et al., 1999; McDonald et al., in press). A recent meta-analysis estimated annual hippocampal reductions to be 4.66% in AD (Barnes et al., 2009). These were also the ROIs showing the largest volume decrease in the AD patients in the present study, with annual rates of decline of 3.75% for hippocampus and 3.79% for amygdala in the AD sample. Of the surfacebased cortical ROIs showing substantial decline, entorhinal, parahippocampal and lateral temporal, as well as inferior parietal, supramarginal and fusiform areas, are all found to be vulnerable in early stages of AD (Du et al., 2007; Fennema-Notestine et al., 2009; McDonald et al., in press; McEvoy et al., 2009; Singh et al., 2006). An important question regards the extent to which atrophy in these areas in healthy elderly is an indication of preclinical AD. We cannot rule out with certainty that degenerative processes associated with early AD caused some of the observed atrophy. However, reductions were also seen in areas relatively less affected by early AD. For instance, large effects were found in inferior, middle and superior frontal cortex. Previous studies of the ADNI sample did not find differences in thickness between single domain amnestic MCI and controls in these areas (Fennema-Notestine et al., 2009), nor did these areas show longitudinal increases in atrophy in MCI participants showing mild functional impairment (CDR-SB of 1 or less) (McDonald et al., in press). Still, a recent study found prefrontal cortical volume to predict dementia (Burgmans et al., 2009a), and AD involves frontal atrophy (Dickerson et al., 2008; Du et al., 2007; Fjell et al., 2008; Singh et al., 2006). An early longitudinal study found that while temporal atrophy was seen in

presymptomatic and mildly affected patients, frontal involvement occurred later in the disease (Scahill et al., 2002). Another study found demented patients to show annual atrophy rates exceeding 0.4% in large areas of the brain, including the frontal cortex (Buckner et al., 2005). In the present study, more atrophy was observed in AD than in healthy controls in almost all ROIs, including prefrontal cortex. However, temporal lobe atrophy was the most prominent in AD, while healthy aging was characterized by comparable rates of atrophy in the temporal lobe and the frontal cortex. The temporal changes seen in the healthy elderly group may be related to instances of preclinical AD, while the frontal changes are more prominent than what would be expected due to AD. Significant decline was also observed in thalamus, accumbens and cerebral and cerebellar white matter; areas that are not among the most vulnerable in early phases of AD (Fennema-Notestine et al., 2009). In the AD patients included in the present study, cerebellum WM was the 39th most vulnerable region, while thalamus was the 22nd and accumbens the 14th. Thus, it is unlikely that all the brain changes found here in the healthy elderly group can be attributed to preclinical AD. We believe that many of the changes observed in healthy aging are caused by processes unrelated to degenerative disease. Consistent with this, a recent study reported that the transition from healthy elderly to MCI was associated with a unique pattern of longitudinal change, including increased ventricular size, and reductions in temporal grey matter, orbitofrontal areas and temporal association cortices, including the hippocampus (Driscoll et al., 2009). The pattern of atrophy over several years in healthy elderly not developing MCI was widespread but did not resemble the pattern associated with MCI (Driscoll et al., 2009). As further follow-up data become available, the hypothesis that the temporal changes seen in the elderly are related to preclinical AD, while the frontal changes are less associated with incipient disease, can be tested. Since it is possible that structural brain changes due to degenerative disease may be manifested long before cognitive symptoms are detectable, follow-up cognitive and clinical data for several years will be invaluable (Burgmans et al., 2009b).

Accelerated atrophy with higher age

Older age was related to increased atrophy and more ventricular expansion. This is consistent with cross-sectional reports of non-linear volumetric changes (Allen et al., 2005; Fjell et al., In press-b; Jernigan and Gamst, 2005; Walhovd et al., 2005; Walhovd et al., in press). However, cross-sectional studies of cortical thickness have mostly reported linear age-changes over the adult life-span (Fjell et al., in press-a; Salat et al., 2004). It is possible that longitudinal designs are more sensitive to accelerated decline in the oldest old, especially if the effects are of moderate strength and evident only in the latest part of the life-span. The correlations between age and rate of change seldom exceeded +/-.30. Interestingly, the strongest correlations between age and rate of atrophy were found in entorhinal cortex and the hippocampus, and significant correlations were also found in isthmus of the cingulate, middle temporal and parahippocampal gyri, all of which are affected in early stages of AD. Since the occurrence of preclinical AD is much higher among 70- and 80 year olds than among 50 and 60 year olds, it is possible that the pattern of accelerated longitudinal atrophy in aging is related to preclinical AD in a subgroup of the participants, even though cognitive symptoms are not yet manifested. This interpretation is in agreement with another recent study where elderly cognitively normal participants showed a pattern of atrophy more similar to that seen in AD than did younger participants, i.e., more temporal lobe atrophy (Davatzikos et al., 2009).

Conclusion

The present results show that widespread changes in brain structure can be found in healthy elderly over only one year, and that atrophy accelerates with advanced age. Some of the atrophy occurred in areas vulnerable to AD and some in areas less characteristic of the disease in early stages. This suggests that the changes are not primarily driven by degenerative processes associated with AD, although it is likely that preclinical changes associated with AD are

superposed on changes due to normal aging in some subjects. Indeed, the pattern of accelerated atrophy with increasing age is probably influenced by occurrence of preclinical AD in a subgroup of the most elderly participants, and this may especially be reflected in the temporal lobe. Nevertheless, this study shows that decreases in brain volume in healthy aging can be observed in almost all brain areas over 1 to 2 years.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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http://www.loni.ucla.edu/ADNI/Data/ADNI_Authorship_List.pdf

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Figure 1. Cortical atrophy in healthy elderly

General linear models were used to test whether rates of atrophy for one and two years were significantly different from zero. The results were thresholded by a conventional criterion for correction for multiple comparisons (false discovery rate < .05) and projected onto a semi-inflated template brain. Blue-cyan areas indicate cortical reduction, while red-yellow areas indicate expansion. As can be seen, significant volume reductions are found across large areas already after one year, and these were especially prominent in temporal and prefrontal areas.



Figure 2. Rate of change in healthy elderly and AD patients

The left panel shows annual longitudinal atrophy in healthy elderly calculated as percentage volume change relative to baseline, while the middle panel shows longitudinal atrophy over two years. Annual change of about 0.5% can be seen across large sections of the brain surface, e.g. in temporal and prefrontal areas. The right panel shows 1 year atrophy in AD patients. The areas of change in the healthy elderly are overlapping with areas most strongly affected in Alzheimer's disease, e.g. temporal lobe cortex. However, areas that are not especially affected in AD, e.g. prefrontal cortex, also undergo large changes in healthy aging. Please note that to allow optimal visualization of regional variation, the color-scale is different for the HE and the AD groups.



Figure 3. Regional rates of a trophy relative to mean atrophy within group for healthy elderly and $\rm AD$

To visualize the areas that showed more than average atrophy and the areas that showed less than average atrophy within each group, mean cortical volume reduction was calculated for each hemisphere in each group. This value was subtracted from the surface maps, yielding a new map highlighting areas that show greater (red) and lesser (blue) rates of atrophy than average for that group. As can be seen, in healthy elderly (left panel) higher than average volume reductions are seen in lateral temporal, inferior parietal, supramarginal and frontal cortex. In contrast, the AD patients show higher than average atrophy mainly in the temporal lobe (middle and inferior temporal gyrus, fusiform gyrus). Please note that to allow optimal visualization of regional variation, the color-scale is different for the HE and the AD groups.





The line graphs show rate of atrophy for the frontal and the temporal regions in healthy elderly and AD patients. The regions are shown below (blue: frontal, red: temporal). In addition to the cortical areas shown, hippocampus and amygdala were included in the temporal region (yellow and cyan 3D figures). ANOVA showed a significant interaction between group and region: While the AD patients showed more atrophy in the temporal (-2.82%, SE = 0.14) than the frontal regions (-1.26%, SE = 0.11), the rate of atrophy between these two regions was comparable in the healthy controls (temporal: -0.56%, SE = 0.07/frontal: -0.38%, SE = 0.07).



Figure 5. Relationships between age and rate of atrophy in healthy elderly Scatterplots for selected ROIs showing the relationship between age and rates of annual volumetric reductions.

Table 1

Sample characteristics for the healthy elderly group

	Baseline	1 year	2 years
n	142	142	89
Sex (females/males)	68/74	68/74	38/ 51
Age	75.6 (59.8–90.2)	76.7 (60.8–91.3)	78.7 (65.1–92.3)
MMSE	29.2 (25-30)	29.1 (24–30)	29.1 (26–30)
CDR sum of boxes	0.03 (0-0.5)	0 (0–0)	0 (0–0)

Table 2

Percentage volume change over 1 and 2 years in healthy elderly n.s. indicates $p \ge .01$

	% change longitudinal	p <	% change longitudinal	p <
	1 year		2 years	
Volumetric ROIs				
Accumbens	-0.57	10^{-8}	-1.40	10^{-19}
Amygdala	-0.81	10^{-9}	-2.02	10^{-13}
BrainStem	-0.31	10^{-5}	-0.86	10^{-12}
Caudate	-0.24	n.s.	-0.62	.005
Cerebellum GM	-0.35	10^{-8}	-0.94	10^{-15}
Cerebellum WM	-0.57	10^{-14}	-1.22	10^{-20}
Cerebral WM	-0.58	10^{-16}	-1.15	10^{-19}
Hippocampus	-0.84	10^{-14}	-1.76	10^{-15}
Pallidum	-0.40	10^{-11}	-0.75	10^{-13}
Putamen	-0.43	10^{-8}	-0.92	10^{-16}
Thalamus	-0.69	10^{-16}	-1.42	10-21
Ventricular ROIs				
Lat vent	4.40	10^{-21}	9.42	10^{-19}
Inf lat vent	5.47	10^{-15}	12.11	10^{-18}
3rd ventricle	3.07	10^{-12}	6.60	10^{-14}
4th ventricle	0.71	n.s.	2.18	10^{-4}
Surface ROIs				
Cingulate, caudal ant	-0.27	0.001	-0.91	10-12
Cingulate, rostral ant	-0.34	10^{-4}	-0.92	10^{-12}
Cingulate, posterior	-0.25	0.001	-0.79	10 ⁻⁷
Cingulate, isthmus	-0.20	0.005	-0.63	10^{-7}
Frontal, superior	-0.31	0.005	-0.93	10^{-11}
Frontal, caudal middle	-0.40	10^{-6}	-0.99	10^{-12}
Frontal, rostral middle	-0.32	10^{-4}	-0.98	10-13
Frontal, pars opercularis	-0.38	10^{-7}	-1.04	10^{-14}
Frontal, pars triangularis	-0.37	10^{-6}	-1.08	10^{-15}
Frontal, pars orbitalis	-0.38	10^{-4}	-1.11	10-13
Frontal, lateral orbital	-0.42	10 ⁻⁶	-1.07	10^{-10}
Frontal, medial orbital	-0.39	10^{-5}	-0.98	10-14
Frontal, pole	-0.59	10^{-5}	-1.30	10^{-11}
Parietal, precentral gyrus	-0.10	n.s.	-0.48	10-4
Parietal, post central gyrus	0.09	n.s.	0.01	n.s.
Parietal, paracentral gyrus	-0.17	n.s.	-0.52	.001
Parietal, superior	-0.18	n.s.	-0.68	10^{-5}
Parietal, inferior	-0.40	10^{-7}	-1.03	10^{-13}

	% change longitudinal	p <	% change longitudinal	p <
	1 year		2 years	
Parietal, supramarginal	-0.40	10^{-6}	-0.98	10^{-10}
Parietal, precuneus	-0.30	10^{-6}	-0.82	10^{-10}
Temporal, parahippocampal	-0.36	10^{-6}	-1.08	10^{-18}
Temporal, entorhinal	-0.55	10^{-7}	-1.20	10^{-11}
Temporal, pole	-0.56	10^{-5}	-1.17	10 ⁻⁹
Temporal, superior	-0.43	10^{-7}	-1.00	10^{-13}
Temporal, middle	-0.47	10^{-8}	-1.25	10^{-16}
Temporal, inferior	-0.47	10 ⁻⁹	-1.18	10^{-17}
Temporal, transverse	-0.31	.005	-0.79	10^{-8}
Temporal, Banks Sup Temp Sulc	-0.55	10^{-11}	-1.32	10 ⁻¹⁵
Temporal, fusiform	-0.38	10 ⁻⁹	-0.98	10^{-18}
Occipital, lateral	-0.19	0.005	-0.66	10 ⁻⁹
Occipital, pericalcarine	0.11	n.s.	-0.18	n.s.
Occipital, lingual	-0.02	n.s.	-0.35	10^{-4}
Occipital, cuneus	-0.04	n.s.	-0.40	10^{-4}

Table 3

Percentage volume change over 1 year in the AD group n.s. indicates $p \geq .01$

	% change longitudinal	p <
	1 year	
Volumetric ROIs		
Accumbens	-2.02	10^{-19}
Amygdala	-3.79	10^{-19}
BrainStem	-0.65	10^{-16}
Caudate	-1.49	10^{-13}
Cerebellum GM	-0.99	10^{-25}
Cerebellum WM	-0.95	10^{-18}
Cerebral WM	-1.42	10^{-30}
Hippocampus	-3.75	10^{-35}
Pallidum	-0.90	10^{-15}
Putamen	-1.45	10^{-22}
Thalamus	-1.51	10^{-28}
Ventricular ROIs		
Lat vent	10.35	10^{-28}
Inf lat vent	16.26	10^{-33}
3rd ventricle	6.82	10^{-21}
4th ventricle	1.61	.001
Surface ROIs		
Cingulate, caudal ant	-1.14	10^{-17}
Cingulate, rostral ant	-1.20	10^{-18}
Cingulate, posterior	-1.57	10 ⁻²¹
Cingulate, isthmus	-1.54	10 ⁻²³
Frontal, superior	-1.20	10^{-15}
Frontal, caudal middle	-1.60	10^{-20}
Frontal, rostral middle	-1.23	10^{-15}
Frontal, pars opercularis	-1.31	10^{-18}
Frontal, pars triangularis	-1.10	10^{-17}
Frontal, pars orbitalis	-1.06	10^{-10}
Frontal, lateral orbital	-1.47	10^{-18}
Frontal, medial orbital	-1.19	10^{-17}
Frontal, pole	-1.38	10^{-10}
Parietal, precentral gyrus	-0.69	10^{-10}
Parietal, post central gyrus	-0.27	.01
Parietal, paracentral gyrus	-0.76	10^{-11}
Parietal, superior	-1.26	10^{-18}
Parietal, inferior	-2.10	10-27

	% change longitudinal	p<
	1 year	
Parietal, supramarginal	-1.74	10^{-21}
Parietal, precuneus	-1.61	10^{-24}
Temporal, parahippocampal	-2.05	10^{-34}
Temporal, entorhinal	-2.42	10^{-36}
Temporal, pole	-2.54	10^{-26}
Temporal, superior	-1.63	10^{-23}
Temporal, middle	-2.87	10^{-31}
Temporal, inferior	-2.97	10^{-32}
Temporal, transverse	-0.59	10^{-6}
Temporal, Banks Sup Temp Sulc	-2.53	10^{-28}
Temporal, fusiform	-2.15	10^{-33}
Occipital, lateral	-0.96	10^{-19}
Occipital, pericalcarine	-0.16	n.s.
Occipital, lingual	-0.67	10^{-15}
Occipital, cuneus	-0.50	10 ⁻⁹

Table 4

Pearson correlations between age and 1-year volume change in healthy elderly Negative correlations between age and rate of atrophy, and positive correlations between age and rate of ventricular expansion are generally found, showing that increasing age is associated with more atrophy and increased ventricular size.

	1 year change
Volumetric ROIs	
Accumbens	-0.13
Amygdala	-0.16
BrainStem	-0.15
Caudate	-0.06
Cerebellum GM	<u>-0.20</u>
Cerebellum WM	<u>-0.17</u>
Cerebral WM	<u>-0.17</u>
Hippocampus	<u>-0.33</u>
Pallidum	<u>-0.17</u>
Putamen	-0.14
Thalamus	<u>-0.18</u>
Ventricular ROIs	
Lateral ventricles	0.08
Inf lateral ventricles	0.21
3rd ventricle	0.03
4th ventricle	0.08
Surface ROIs	
Cingulate, caudal ant	<u>-0.19</u>
Cingulate, rostral ant	-0.11
Cingulate, posterior	-0.22
Cingulate, isthmus	-0.25
Frontal, superior	-0.13
Frontal, caudal middle	-0.21
Frontal, rostral middle	-0.13
Frontal, pars opercularis	-0.26
Frontal, pars triangularis	-0.23
Frontal, pars orbitalis	-0.08
Frontal, lateral orbital	<u>-0.19</u>
Frontal, medial orbital	<u>-0.18</u>
Frontal, pole	-0.11
Parietal, precentral gyrus	-0.21
Parietal, post central gyrus	<u>-0.19</u>
Parietal, paracentral gyrus	-0.13
Parietal, superior	<u>-0.19</u>
Parietal, inferior	<u>-0.20</u>
Parietal, supramarginal	<u>-0.19</u>
Parietal, precuneus	<u>-0.20</u>

	1 year change
Temporal, parahippocampal	-0.23
Temporal, entorhinal	<u>-0.32</u>
Temporal, pole	<u>-0.20</u>
Temporal, superior	-0.22
Temporal, middle	-0.23
Temporal, inferior	<u>-0.17</u>
Temporal, transverse	<u>-0.30</u>
Temporal, Banks Sup Temp Sulc	-0.23
Temporal, fusiform	<u>-0.20</u>
Occipital, lateral	<u>-0.17</u>
Occipital, pericalcarine	<u>-0.18</u>
Occipital, lingual	-0.16
Occipital, cuneus	-0.13

 $\underline{\textit{Underlined italic}}$ indicates $p < .05 \ (r \geq +/- \ .17)$

Bold indicates $p < .01 \ (r \geq +\!/\!-.21)$

<u>Bold underlined italic</u> indicates p < .001 ($r \ge +/-.27$)